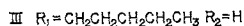
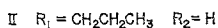
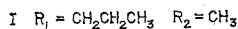
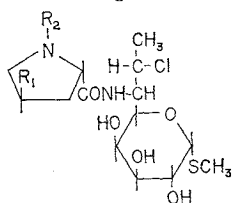


MICROBIAL TRANSFORMATION
OF ANTIBIOTICS VII.
HYDROXYMETHYLATION OF
N-DEMETHYLCLINDAMYCIN

Sir:

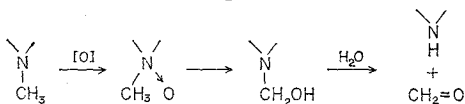
We have reported¹⁾ that several streptomycetes transform clindamycin (I, Fig. 1) to N-demethylclindamycin (II, Fig. 1).

Fig. 1.



The mechanism of this enzymatic demethylation is not known. However, enzymatic N-demethylation of tertiary amines is believed^{2,3)} to proceed by the steps outlined in Fig. 2.

Fig. 2.

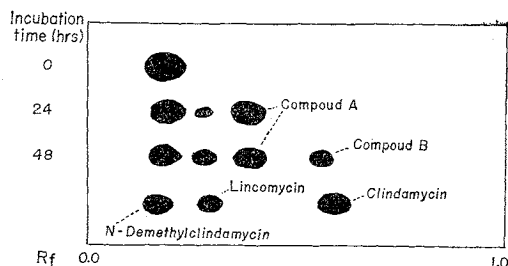


We were interested in determining whether the methylation of N-demethylclindamycin to clindamycin could also be carried out by streptomycetes. We selected *Streptomyces lincolnensis* for these studies, since this organism contains⁴⁾ efficient methylating systems.

N-Demethylclindamycin was added to 24-hour cultures of *S. lincolnensis*⁵⁾ at levels of 50~100 mcg/ml. As shown in Fig. 3, immediately after the addition of N-demethylclindamycin (0 time) this antibiotic was the only bioactive compound in the fermentation broth. Twenty four hours after addition of the compound, part of the N-demethylclindamycin had been transformed to a new bioactive material, compound A. At that time the fermentation broth also contained lincomycin normally produced by the culture. Forty eight hours after addition, tlc showed

Fig. 3. Thin-layer chromatography* of fermentations of *Streptomyces lincolnensis* containing N-demethylclindamycin.

* Silica Gel G: Ethyl acetate-acetone-water (8:5:1 v/v). Antibiotics were detected by bioautography on *S. lutea*-seeded agar.



the presence of N-demethylclindamycin, lincomycin, compound A, and a new bioactive material, compound B. *S. lincolnensis* growing in the absence of N-demethylclindamycin, produced lincomycin only. Since less compound A was present in the 48-hour sample than in the 24-hour sample it was assumed that both compounds A and B are biotransformation products of N-demethylclindamycin and that A is formed first and slowly transformed to compound B.

4'-Depropyl-4'-pentyl-N-demethylclindamycin (III, Fig. 1) was also transformed by *S. lincolnensis*. Clindamycin, however, was not effected at all by this organism. This suggested the imino group of propyl proline moiety of N-demethylclindamycin as the site of biotransformation.

We directed our efforts to the isolation of compound A since this material was the main bioconversion product of N-demethylclindamycin. The activities present in the fermentation broth were adsorbed on Amberlite XAD-2 and eluted using 95% aqueous methanol. Compound A is very unstable and is converted to N-demethylclindamycin and small amounts of compound B when subjected to countercurrent distribution, silica gel, and partition or Sephadex chromatographies. When CM-Sephadex, a cationic exchanger was used, N-demethylclindamycin and lincomycin were adsorbed, while compound A passed through the column. This compound, however, was partially converted to N-demethylclindamycin and compound B during the recovery process. The behavior of compound A suggested a neutral or weakly basic compound which

Fig. 4.

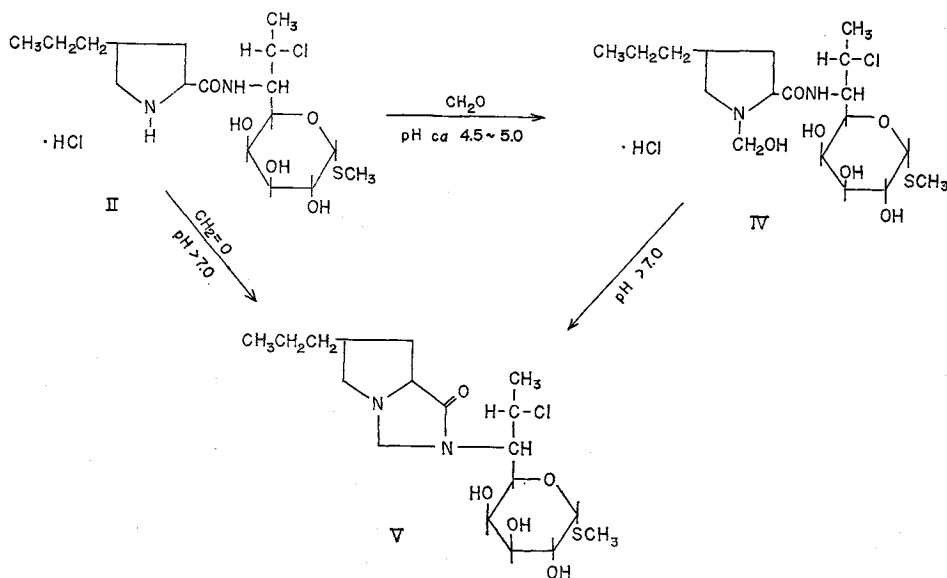
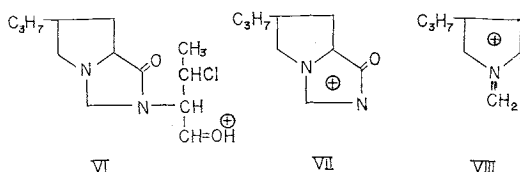


Fig. 5.



could be degraded to N-demethylclindamycin under mild conditions. N-Demethyl-N-hydroxymethylclindamycin (IV, Fig. 4), is such a compound. This material is expected to be unstable and to be easily degraded to N-demethylclindamycin and formaldehyde. One also would expect that hydroxymethylation of the secondary amino-group would decrease the basic properties of N-demethylclindamycin⁹. This is in agreement with the fact that compound A was not adsorbed by CM-Sephadex. Compound A under isolation conditions was transformed not only to N-demethylclindamycin but to compound B. Therefore, any structure proposed for compound A should take into account this transformation.

We proceeded to synthesize N-demethyl-N-hydroxymethylclindamycin. Mixing an aqueous solution of N-demethylclindamycin hydrochloride with formalin at room temperature gave a compound with Rf values greater than those of N-demethylclindamycin and identical to those of compound A. The

product of the reaction, assigned structure IV, was isolated as a colorless, amorphous, highly hygroscopic material, showing secondary amide carbonyl absorption at 1680, and 1500 cm^{-1} in the IR spectrum. This material was highly unstable and decomposed to N-demethylclindamycin and formaldehyde. Mass spectra were not helpful in establishing the structure. The NMR spectrum on the other hand, shows the presence of all the groupings present in N-demethylclindamycin and in addition an absorption peak at $\text{ca } \delta, 5.2$ assigned to the N-hydroxymethyl group.

When the reaction of N-demethylclindamycin and formaldehyde was carried out at pH greater than 7.0, the product of the reaction was a new bioactive compound which had Rf values identical to those of compound B. In addition when solutions of N-demethyl-N-hydroxymethylclindamycin were adjusted to pH greater than 7.0 the new compound was formed almost quantitatively. This compound, isolated crystalline, was found to be very stable and to have structure V (Fig. 4).

Analytical data agree with the molecular formula of $\text{C}_{13}\text{H}_{21}\text{ClN}_2\text{O}_5\text{S}$ for V. Potentiometric titration in water shows the presence of one weakly basic group with pK_a' of 4.1 (Eq. weight found, 418). The IR spectrum shows the absence of the amide II

band (at ca 1,500 cm^{-1}) common to all linco-saminides indicating a tertiary amide carbonyl. The mass spectrum contains molecular ion peak at m/e 422,1616. (Theor. mol. weight, 422,1642). The spectrum also shows peaks at M^+-SCH_3 , M^+-H_2O , M^+-HCl and M^+-CH_3 . Large peaks at m/e 273, 163 and 125 are assigned to ion fragments VI, VII, VIII (Fig. 5), respectively.

Silylation of V yielded a compound which showed molecular ion peak at 638 mass units indicating the presence of three trimethylsilyl groups in the molecule. These data establish the bicyclic imidazolidone structure V for the crystalline product of the reaction of N-demethylclindamycin and formaldehyde at elevated pH.

As mentioned earlier, compounds A and B, produced when N-demethylclindamycin was added to fermentations of *S. lincolnensis*, were found to have tlc mobilities identical to those of N-demethyl-N-hydroxymethylclindamycin (IV) and the bicyclic derivative (V), respectively. There is little doubt that compound A is N-demethyl-N-hydroxymethylclindamycin and that this material is produced by microbial hydroxymethylation. Compound B on the other hand, identical to the bicyclic imidazolidone V, probably is formed from compound A by dehydration under the fermentation conditions.

The biological properties of N-demethyl-N-hydroxymethylclindamycin and imidazolidone V will be reported in a subsequent communication.

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